# GROWTH, CELL WALL ELASTICITY AND PLASTICITY IN ZEA MAYS L. COLEOPTILES EXPOSED TO CADMIUM

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#### RESUMEN

# CRECIMIENTO Y PROPIEDADES ELASTICAS Y PLASTICAS DE LAS PAREDES CELULARES DE COLEOPTILOS DE ZEA MAYS L. EXPUESTOS A CADMIO

Se ha estudiado la influencia de diferentes concentraciones de Cd (0, 0.05, 0.1, 1 y 10 mM) sobre el crecimiento en extensión y las propiedades físicas de las paredes celulares de coleoptilos de *Zea mays* L. El crecimiento en extensión fué seguido de forma continuada mediante un sistema de transductor de desplazamiento computerizado. El mismo sistema modificado permitió el análisis de las propiedades físicas de las paredes celulares, estimándose la extensibilidad total, la elasticidad y la plasticidad mediante la técnica Instron modificada. Todas las concentraciones de Cd suministradas inhibían de forma significativa el crecimiento. No se observó una correlación clara entre las propiedades de extensibilidad total, plasticidad y elasticidad de las paredes celulares con las concentraciones de Cd suministradas. De acuerdo con nuestros resultados la alteración de las propiedades físicas de las paredes celulares no parece ser la causa inicial de la disminución del crecimiento en extensión de coleoptilos sometidos a toxicidad por Cd.

Palabras clave: Cadmio. Inhibición crecimiento. Paredes celulares. Extensibilidad. Plasticidad. Sistema computerizado de desplazamiento lineal.

#### SUMMARY

The influence of different Cd concentrations (0,0.05,0.1, 1 and 10 mM) on extension growth and physical properties of cell walls was studied in *Zea mays* L. coleoptiles. Extension growth of the coleoptiles was continuously monitored with a computerized linear displacement transducer system. The same system, somewhat modified, was used for the analysis of the physical properties of the coleoptile cell walls. Total extensibility, cell wall plasticity and elasticity was measured by a modified Instron technique. All Cd concentrations supplied significantly inhibited extension growth of the coleoptiles. No clear correlation between the Cd concentrations supplied and cell wall extensibility, plasticity and elasticity was observed. We may conclude from our results that alteration of physical cell wall properties was not a primary cause for inhibition of extension growth by Cd in maize coleoptiles.

Key words: Cadmium. Growth inhibition. Cell wall. Extensibility. Plasticity. Elasticity. Computerized linear displacement transducer system.

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#### INTRODUCTION

Cell expansion growth generally is described by the modified Lockhart's equation (Dale and Sutcliffe, 1986):

$$\frac{\mathrm{dV}}{\mathrm{dt}} = \frac{\mathrm{L'}_{p}\phi \left(\Delta\Psi + \mathrm{P} - \mathrm{Y}\right)}{\phi + \mathrm{L'}_{p}}$$

where  $L_p$  is the apparent hydraulic conductance,  $\phi$  the extensibility of the wall,  $\Delta\Psi$  the difference of water potential between the cell and its surroundings, Y the threshold turgor pressure, and P the actual turgor pressure. Any change of these parameters would alter the rate of cell expansion growth.

Cell expansion growth is generally severely inhibited by metal toxicity (Barceló and Poschenrieder, 1990; Breckle, 1991). It has been suggested that toxic levels of metals such as Al, Cd, Zn or Pb may bind to the pectin fraction of cell walls (Klimashevski and Dedov, 1975; Khan et al., 1984; Peterson, 1969; Wierzbicka, 1987) and inhibit cell expansion growth by affecting cell wall extensibility. Cross-linking of the pectin carboxyl groups by metals has been suggested as a possible mechanism, but it seems more likely that metals affect physical properties of cell walls by influencing the synthesis and assambly of cell wall components. (Barceló and Poschenrieder, 1990; Barceló, 1991).

In the present short-term study we analysed the influence of different Cd concentrations on growth, cell wall extensibility, plasticity and elasticity, in order to establish the possible role of Cd-induced alterations of these parameters in growth inhibition by Cd.

### MATERIALS AND METHODS

Two types of experiments were performed. *In vivo* measurement of extension growth (experiment I) and determination of *In-vitro* extensibilities of cell walls (experiment II).

Experiment I: For investigating the influence of Cd on the extension growth of maize coleoptiles, Zea mays L. cv Honeycomb (Semillas Batlle SA, Barcelona) seeds were germinated for seven days on sterilized Petri dishes in darkness (25 °C). Only uniform seedlings with stright coleoptiles of an initial length of approximately 15 mm were used.

For extension growth monitoring, a linear displacement transducer de-

vice (LVDT) similar to that described by Penny et al. (1973) and modified by Cramer et al. (1988) was used. The transducer (Sangamo, U.K., type DF-1 with a DCU1B conditioner) was connected to the coleoptile with a nylon thread. Data adquisition was made with a personal computer (PC/XT, Elbe Microsystems, PCX, Barcelona). Seedlings were grown in continuously aerated, flowing solutions, containing 0.4 mM Ca (controls) or 0.05, 0.1, 1 or 10 mM Cd as CdCl<sub>2</sub>.2.5 H<sub>2</sub>O). A scheme of our device is shown in figure 1.

Growth of the coleoptiles was monitored for 4 h. After this, roots

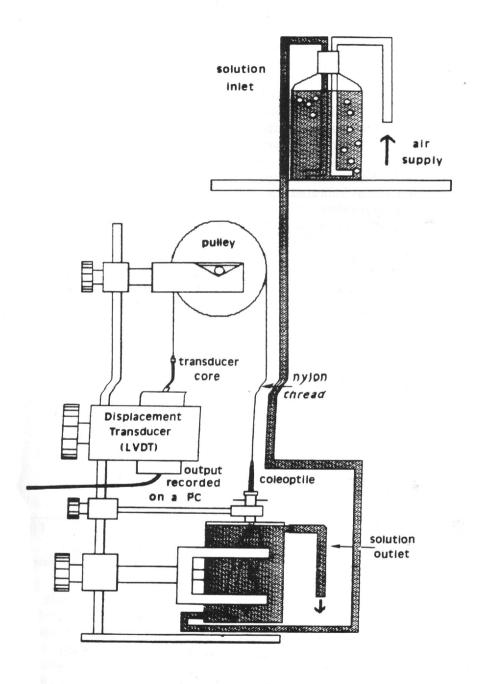


FIG. 1.—Device for extension growth monitoring (modified after Penny et al., 1973).

were submerged for 30 min in 0.4 mM  $CaCl_2$  solution, followed by distilled water. The plant material was dried at 60  $^{\circ}C$  and, after digestion (0.1 g dry matter in 0.2 mL  $HClO_4$  and 1.4 mL  $H_2O_2$  in closed vials), Ca and Cd concentrations were determined by atomic absorption spectrometry (Perkin Elmer 703).

Experiment II: Seeds of Zea mays L. were germinated as indicated above. Ten uniform coleoptiles per treatment were cut and placed in Petri dishes with 15 mL of control or Cd-containing solution (0.05, 0.1 1 or 10 mM as CdCl<sub>2</sub>.2H<sub>2</sub>O). The dishes were gently shaken for 2, 4 or 24 h. After washing with a 0.4 mM Ca solution, the coleoptiles were

frozen at -30 °C. Before measuring cell wall extensibility, coleoptiles were allowed to thaw.

Cell wall extensibility, elasticity and plasticity were determined with the Instron technique according to Kutschera y Schopfer (1986). The same linear displacement transducer from experiment I was used. The weight for cell wall extension was 20 g. The displacement caused by the cell wall extension was monitored by a personal computer as indicated above. Total extensibility (Et), elastic extensibility (Ep) were determined on the graphics as shown in figure 2.

Cadmium and Ca concentrations of coleoptiles were determined as in experiment I.

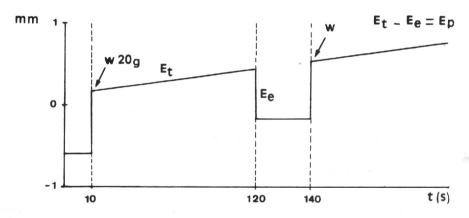


FIG. 2.—Schematic representation of an in vitro extensibility curve, indicating total extensibility, plastic and elastic extensibility (Kutschera and Schopfer, 1985).

### RESULTS

In vivo growth measurement showed that all Cd concentrations assayed had a significant inhibitory effect on coleoptile growth (Fig. 3).

The growth rate of control coleoptiles generally increased during the whole experiment (4 h) (Fig. 4). The growth rates of coleoptiles exposed

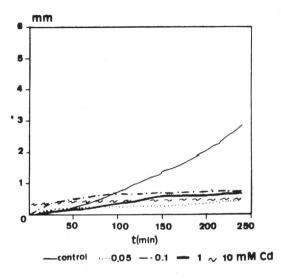


FIG. 3.—Growth (mm) of Zea mays L. coleoptiles grown in control or Cd-containing nutrient solution.

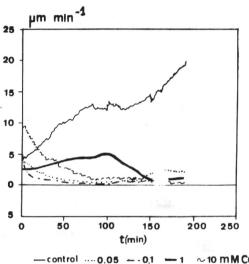


FIG. 4.—Growth rates (µm min<sup>-1</sup>) Zea mays L. coleoptiles grown in control or Cd-containing nutrient solution.

to Cd generally decreased inmediately after Cd supply and felt to zero after approximately 90 min exposure to Cd. Coleoptiles exposed to 1 mM Cd showed a different behaviour, reaching a zero growth rate after approximately 150 min exposure to Cd. Coleoptiles exposed to 0.1 mM Cd or higher concentrations did not recover a significant growth rate

within the experimental time, while coleoptiles treated with 0.05 mM exhibited a small increase of growth rate after approximately 145 min.

Table 1 shows the concentrations of Ca and Cd in roots and coleoptiles. All plants treated with Cd showed higher Ca concentrations in roots and lower Ca concentrations in shoots than controls. The lowest Ca

|                            | TABLE 1   |                                  |
|----------------------------|---|----------------------------------|
| Concentration of Ca and Cd | (μg g <sup>-1</sup> ) in roots and coleops<br>or Cd-Containing solutions. | tiles exposed for 4 h to control |

| Cd conc. in solution | Ro      | Roots   |         | Coleoptiles |  |
|----------------------|---------|---------|---------|-------------|--|
|                      | Calcium | Cadmium | Calcium | Cadmium     |  |
| Control              | 10.68   | n. d.*  | 15.24   | n, d.       |  |
| 0.05 mM              | 18.95   | 166.67  | 8.94    | n.d.        |  |
| 0.1 mM               | 17.26   | 283.43  | 4.99    | n.d.        |  |
| 1 mM                 | 15.34   | 1324.17 | 10.40   | 7.97        |  |
| 10 mM                | Missing | Missing | 12.31   | 307.12      |  |

<sup>\*</sup> n. d., not detectable.

concentrations were found in shoots of the 0.1 mM Cd treatment. The Cd concentrations within both roots and shoots significantly increased with Cd supply. In control plants and in shoots of plants exposed to 0.05 or

0.1 mM Cd, the Cd concentrations were below the detection limit.

Table 2 shows the Ca and Cd concentrations of coleoptiles floated for different times (2 h, 4 h or 24 h) on control (0.4 mM Ca) or Cd-containing

TABLE 2

Calcium and Cadmium concentrations of coleoptiles floated for different times (2 h, 4 h, 24 h) on control (0.4 mM Ca) or Cd-containing solutions.

| EXPOSURE   | CADMIUM CONCENTRATION IN SOLUTION |         |         |                |         |
|--|-----------------------------------|---------|---------|----------------|---------|
| TIME   | Control                           | 0.05 mM | 0.1 mM  | 1.0 m <b>M</b> | 10 mM   |
| Calcium concentration $\mu_g$ g <sup>-1</sup> dry weight |                                   |         |         |                |         |
| 2 h  | 8.11                              | 12.53   | 17.60   | 39.10          | 1.04    |
| 4 h  | 23.83                             | 3.74    | 16.64   | 12.49          | 122.48  |
| 24 h   | 18.73                             | 31.75   | 225.55  | 17.08          | 20.12   |
| Cadmium concentration $\mu$ g g <sup>-1</sup> dry weight |                                   |         |         |                |         |
| 2 h  | n. d.*                            | n. d.   | 107.40  | 625.61         | 1009.52 |
| 4 h  | n. d.                             | 147.87  | Missing | 893.37         | 638.21  |
| 24 h   | n.d.                              | 286.71  | 743.49  | 3159.29        | 9498.58 |

<sup>\*</sup> n. d., not detectable.

solutions. Generally both Cd treatment and exposure time significantly influenced Ca concentrations of coleoptiles, but no correlation between Cd supply and Ca concentration could be established. The cadmium concentration in the coleoptiles significantly increased with the Cd concentration supplied and with the exposure time.

Total extensibility, elastic and plastic extensibility of cell walls are shown in Table 3. In controls, a significant decrease with time of total extensibility and plastic and elastic extensibility was observed. In coleoptiles exposed to Cd the response

of physical cell wall properties was more complex. Generally an increase of total extensibility was observed after 2 or 4 h exposure. After 24 h exposure all Cd-treated coleoptiles showed a higher total extensibility than controls. A similar behaviour was found for plastic extensibility. In control plants, the elastic extensibility significantly decreased with time. After 2 h exposure, plants treated with Cd showed significantly lower cell wall elasticity than controls, but differences were not significant after 4 h exposure. After 24 h, coleoptiles treated with 0.05 mM Cd showed significantly higher, and

TABLE 3

Total extensibility, plastic extensibility and elastic extensibility (µm) of cell walls of maize coleoptiles exposed for different times to control or Cd-containing solutions.

| EXPOSURE                       | TOTAL EXTENSIBILITY             |                                   |                                   |                                    |                                  |
|--------------------------------|---------------------------------|-----------------------------------|-----------------------------------|------------------------------------|----------------------------------|
| TIME                           | Control                         | 0.05 mM                           | 0.1 mM                            | 1.0 mM                             | 10 mM                            |
| 2 hours<br>4 hours<br>24 hours | 608.89ae*<br>401.17b<br>327.76b | 423.75bc<br>689.25bc<br>458.15cdf | 640.17ace<br>612.91c<br>401.38bdf | 489.82ce<br>632.34c<br>417.02bcf   | 862.70e<br>642.68ce<br>545.60ef  |
| 2                              |                                 | PLASTIC EXTENSIBILITY             |                                   |                                    | 343,0001                         |
|                                | Control                         | 0.05 mM                           | 0.1 mM                            | 1.0 mM                             | 10 mM                            |
| 2 hours<br>4 hours<br>24 hours | 301.51a<br>290.69ad<br>183.11b  | 244.43bd<br>529.64de<br>246.16bd  | 393.76abc<br>678.86be<br>349.41bf | 331.65abc<br>464.85cde<br>306.40cf | 626.27ce<br>467.92de<br>386.60ce |
|                                | ELASTIC EXTENSIBILITY           |                                   |                                   |                                    |                                  |
|                                | Control                         | 0.05 mM                           | 0.1 mM                            | 1.0 mM                             | 10 mM                            |
| 2 hours<br>4 hours<br>24 hours | 307.38a<br>110.48bd<br>144.65bc | 179.32be<br>168.62bde<br>211.99e  | 246.41ab<br>134.04d<br>51.97f     | 158.17b<br>167.49bd<br>110.61bfg   | 236.43b<br>174.76bd<br>158.93ceg |

<sup>\*</sup> Values followed by the same letter are not significantly different (p < 0.05).

those treated with 0.1 mM Cd significantly lower cell wall eslasticity than controls. No significant corre-

lation between Cd concentration and cell wall extensibility could de established.

## DISCUSSION

Our results confirm earlier findings, that Cd causes severe decrease of extension growth in plants (Poschenrieder et al., 1989). Our observation that extension growth of coleoptiles decreased almost immediately after supply to roots of Cd concentrations which were not translocated in detectable amounts to the coleoptiles (0.05 mM and 0.1 mM Cd treatments), suggests that inhibition of extension growth in the above-ground parts was not due to an in situ effect of Cd, but a consequence of the translocation of a root-induced signal. In plants exposed for 4 h to 1 mM Cd a significant amount of Cd was detectable in the coleoptile, but growth rate was less affected than that of plants receiving lower Cd supply.

Our results on *in vitro* extensibility of coleoptiles floated on control solutions are in line with the hypothesis that cell wall extensibility

and elasticity decrease with plant age and increase of cell size (Tyree and Jarvis, 1982). The tissue Cd concentrations achieved by floating sectioned coleoptiles on Cd solutions were much higher than toxic tissue concentrations reported for maize plants (Bingham et al., 1975). Nevertheless. Cd did not decrease but increase total extensibility of cell walls, which may be related to the smaller cell size in Cd-treated plants. In coleoptiles exposed to Cd, we only observed a transient decrease of elastic extensibility after 2 h exposure. Excepting coleoptiles from the 0.1 mM Cd treatment, values similar or even higher than that of controls were achieved after 24 h. Thus the short-term effect of Cd on cell wall elasticity in maize was clearly distinct from the long-term Cd effect described in Phaseolus vulgaris L. leaves (Barceló et al., 1986).

### CONCLUSIONS

We may conclude from our results, that the Cd-induced inhibition of extension growth in coleoptiles of Zea mays L. is initially not caused by a decrease of cell wall extensibi-

lity. Toxic effects in roots seem to be responsible for the initial growth inhibitory response in upper plant parts.

#### REFERENCES

- BARCELO, J., 1991. Home-plata-medi ambient: una hamonia trencada. Discurso Ingreso Real Academia de Farmacia de Barcelona. Publicaciones Universidad Autónoma de Barcelona, Bellaterra.
- BARCELO, J. and POSCHENRIEDER, CH., 1990. Plant water relations as affected by heavy metal stress: a review, J. Plant Nutr., 13: 1-37.
- BARCELO, J., POSCHENRIEDER, CH., ANDREU, I. and GUNSE, B., 1986. Cadmium-induced decrease of water stress resistance in bush bean plants (*Phaseolus vulgaris* L. cv Contender). I. Effects of Cd on water potential, relative water content and cell wall elasticity. J. Plant Physiol., 125: 17-26.
- BINGHAM, F. T., MAHLER, R. J. and GANJE, T. J., 1975. Growth and cadmium accumulation of plants grown on soil treated with cadmium enriched sewage sludge. J. Environ. Qual., 4: 207-210.
- BRECKLE, S., 1991. Growth under stress: heavy metals. In: Plant roots: the hidden half. Y. Waisel, A. Eshel and U. Kafkafi (Eds.). 351-373. Marcel Dekker, Inc., New York.
- CRAMER, G. R., EPSTEIN, E. and LAUCHLI, A., 1988. Kinetics of root elongation of maize in response to short-term exposure to NaCl and elevated calcium concentrations, J. Exp. Bot., 39: 1513-1522.
- DALE, J. E. and SUTCLIFFE, J. F., 1986. Water relations of plant cells. In: Plant Physiology, a treatise., 9. Water and solutes in plants. F. C. Steward (Ed.). 1-48. Academic Press, Inc., London.
- KHAN, D. H., DUCKETT, J. G., FRANKLAND, B. and KIRKHAM, J. B., 1984. An X-ray analytical study of the distribution of cadmium in roots of *Zea mays* L. J. Plant Physiol., 115: 19-28.
- KLIMASHEVSKI, E. L. and DEDOV, V. M., 1975. Localization of the mechanism of growth inhibiting action of Al<sup>3+</sup> in elongating cell walls. Fiziol. Rast., 22: 1040-1046.
- KUTSCHERA, V. and SCHOPFER, P., 1985. Evidence against the acid growth theory of auxin action, Planta, 163: 483-493.
- PENNY, D., PENNY, P. and MARSHALL, D. C., 1973. High resolution measurement of plant growth. Can. J. Bot., 52: 959-969.
- PETERSON, P. J., 1969. The distribution of zinc-65 in Agrostis tenuis Sibth. and A. stolonifera tissues. J. Exp. Bot., 20: 863-875.
- POSCHENRIEDER, CH., GUNSE, B. and BARCELO, J., 1989. Influence of cadmium on water relations, stomatal resistance, and abscisic acid content in expanding bean leaves. Plant Physiology, 90: 1365-1371.
- TYREE, M. T. and JARVIS, P. G., 1982. Water in tissues and cells. In: Physiological Plant Ecology II. Water relations and carbon assimilation. Encyclopedia of Plant Physiology, New Series. 12 B. O. L. Lange, P. S. Nobel, C. B. Osmond and H. Ziegler (Eds.), 35-77. Springer Verlag, Berlin.
- WIERZBICKA, M., 1987. Lead accumulation and its translocation barriers in roots of Allium cepa L.—autoradiographic and ultrastructural studies. Plant Cell and Environment, 10: 17-26.

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